PALEOECOLOGY OF COMMENSAL EPIZOANS FOULING FLEXICALYMENE (TRILOBITA) FROM THE UPPER ORDOVICIAN, CINCINNATI ARCH REGION, USA

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ABSTRACT—Commensal epizoozoans and episkeletozoans are rarely preserved attached to the external exoskeleton of the Late Ordovician trilobite *Flexicalymene*. Of nearly 15,000 *Flexicalymene* specimens examined, 0.1% show epizoozoans or episkeletozoans. Factors limiting *Flexicalymene* fouling include a shallow burrowing life style, frequent molting of the host, larval preference for other substrates, observational bias caused by overlooking small fouling organisms, and the loss of the non-calcified, outermost cuticle prior to fossilization or as the trilobite weathers from the encasing sediment. Trepostome bryozoans, articulate and inarticulate brachiopods, cornulitids, and a tube-dwelling/boring nonbiomineralized organism represent the preserved members of the Late Ordovician marine hard substrate community fouling *Flexicalymene*. This assemblage of organisms is less diverse than the hard substrate community fouling Late Ordovician sessile epibenthic organisms. Fouling is not restricted to only large *Flexicalymene* specimens as observed in previous studies but occurs in medium to large individuals interpreted as early to late holaspid specimens.

Epizoozoans fouling the carcasses or molt ensembles of 16 *Flexicalymene* specimens provide insight into the life habits of the host and these fouling organisms. Trepostome bryozoans, articulate and inarticulate brachiopods, and cornulitids preferentially attached to elevated portions of the dorsal exoskeleton, and preferentially aligned in either the direct line or lee side of currents generated by *Flexicalymene* walking on the sea floor or swimming through the water column.

INTRODUCTION

F OSSIL AND modern marine hard substrate communities contain a diverse assemblage of organisms. Among them are algae, annelids, barnacles, brachiopods, bryozoans, coccoliths, echinoderms, foraminiferans, hydrozoans, mollusks, serpulid polychaetes, poriferans, protoctists, ascidian tunicates, rugose and tabulate corals, cornulitids, and various echinoderms such as edrioasteroids, cystoids, and blastoids (Taylor and Wilson, 2003). Larvae of these organisms colonize rocks, plants, and the skeletons of living and dead animals by cementing themselves to surfaces or boring into them (Taylor and Wilson, 2002). Larvae are more likely to either settle on sessile epibenthic hosts than on motile benthic or nektonic hosts, or to cement themselves to previously settled members of the hard substrate community (Taylor, 1990). Motile benthic and nektonic mollusks, arthropods, echinoderms, and vertebrates are less frequent sites of colonization (Key et al., 1995, 1996a, 1996b, 1997). Lower colonization rates exhibited by motile hosts may reflect host behaviors developed to remove fouling organisms. For example, modern arthropods can molt their exoskeletons or burrow into the sediment to cleanse themselves of epibionts (e.g., Ross, 1983; Mikulic, 1990; Gili et al., 1993).

In this study, we investigate organisms of the marine hard substrate community that have fouled carcasses and articulated exuviae of the calymenid trilobite *Flexicalymene* spp. Shirley, 1936, which occurs throughout Upper Ordovician rocks of the Cincinnati arch region of southwestern Ohio, southeastern Indiana, and northern Kentucky (Hunda and Hughes, 2007). This is a period when the evolution of epibiotic relationships played an important role in the Great Ordovician Biodiversification Event (Servais et al., 2009). We restrict our investigation to those organisms that cemented themselves to the external skeletons of living or dead *Flexicalymene* spp. or that live within bryozoan colonies fouling *Flexicalymene* spp. We apply the nomenclature for marine organism-hard substrate relationships proposed by Taylor and Wilson (2002). An organism interpreted as colonizing a living *Flexicalymene* spp. host is termed an epizoozoan, and an organism living on a *Flexicalymene* spp. skeleton, whether it belonged to a living animal or a corpse, is termed an episkeletozoan.

Fossil episkeletozoans are valuable resources in the paleoecologic study of ancient hard substrate communities and provide vital information for addressing the life habits of host organisms (Alexander and Brett, 1990; Powers and Ausich, 1990; Lescinsky, 1996; Taylor and Wilson, 2002). Ecologic data commonly lost by preservational bias and destructive taphonomic processes is preserved because fouling episkeletozoans are fossilized in life position on the exoskeletons of host organisms. Thus, hard substrate communities provide opportunities to study the composition, abundance, density, intraspecific spacing, species interactions, larval settling behavior, and overgrowth interactions of its members (Taylor and Wilson, 2003). Examination of episkeletozoan-host interactions also provides information about feeding habits of episkeletozoans and host organisms, the substratum selection of episkeletozoans, and strategies used by living hosts to remove epizoozoans.

Well-preserved trilobite carcasses and articulated exuviae are common fossils in many Paleozoic marine stratigraphic sequences. However, studies describing episkeletozoans on trilobites are relatively few in number (e.g., Brandt, 1996; Kloc, 1997). Moreover, most studies involve few encrusted trilobites (e.g., Prokop, 1965; Kesling and Chilman, 1975; Clarkson and Tripp, 1982; Snajdr, 1983; Mikulic, 1990; Tetreault, 1992, 1997; Budil and Saric, 1995; Kácha and Šaric, 1995; Taylor and Brett, 1996; Brett and Taylor, 1997; Pratt, 2001). Exceptions are Kloc (1992, 1993, 1997) and Brandt (1996) who documented the frequency, stratigraphic occurrence, and paleoecology of trilobite episkeletozoans from sizeable collections of articulated specimens. Kácha and Šaric's (2009) recent study examined a large population of trilobites, but they were all incomplete, disarticulated and transported exoskeletons.

Kloc (1992, 1993, 1997) discovered that 14 (100%) carefully prepared articulated specimens of the Lower Devonian odontopleurine trilobite Dicranurus hamatus elegantus Campbell, 1977 from Oklahoma contained a diverse assemblage of encrusting bryozoans, brachiopods, corals, foraminifera, worm tubes, and unknown nonbiomineralized organisms. Episkeletozoans were present on trilobites of varying holaspid sizes. Episkeletozoans generally encrusted the cephalon, genal, pleural, and occipital spines more than they encrusted the thorax or pygidium. Kloc (1992, 1993, 1997) suggested that a mutualistic relationship existed between the host and the episkeletozoans. D. hamatus elegantus provided the necessary hard substrate as well as increased feeding opportunities and mobility for the sessile episkeletozoans, whereas the episkeletozoans produced a living camouflage that functioned to make the outline of the host less conspicuous to predators.

Tetreault (1992, 1997) examined the autecology of several trilobite species from the Rochester Shale (Silurian) of Ontario and New York. Numerous specimens of the large lichid Arctinurus boltoni (Bigsby, 1825) supported Cornulites sp. Schlotheim, 1820 tubes on the doublure and brachiopods, cyclostome bryozoans, and Cornulites sp. tubes elsewhere on the dorsal exoskeleton. Apertures of Cornulites sp. on the doublure were oriented posteriorly, suggesting a rheophilic response to persistent water circulation patterns beneath the trilobite. Tetreault (1992, 1997) concluded that the upper part of the exoskeleton of Arctinurus sp. remained mud free because of the large number of epizoozoans fouling that area. Further support for the interpretation was provided by the presence of individuals representing several age classes of the brachiopod Stegerhynchus sp. Foerste, 1909a. Tetreault (1992, 1997) concluded that molting had either slowed or stopped in these large holaspid specimens of A. boltoni allowing sufficient time for colonization by epizoozoans.

Brandt (1996) reported that only 8 (0.4%) of 2,030 *Flexicalymene* specimens in museum collections from the Upper Ordovician rocks of the Cincinnati, Ohio, area were fouled with trepostome bryozoans and cornulitid worm tubes. These episkeletozoans preferentially attached to the elevated areas of the anterior/medial portion of the cephalic doublure and the glabella, the axial rings of the thorax, and the medial portion of the pygidium. However, the bryozoans and cornulitids did not exhibit a preferred orientation on the fouled trilobites. Only large *Flexicalymene* spp. specimens interpreted as late adult (holaspid) molt stages contained epizoozoans and episkeletozoans. Brandt (1996) suggested that a commensal relationship existed between *Flexicalymene* and associated episkeletozoans. As interpreted, the exoskeleton of *Flexicalymene* spp. served as a hard substrate for

bryozoans and cornulitids that filtered food from the feeding currents of the host. *Flexicalymene* spp. was interpreted by Brandt (1996) as a semi-infaunal, burrowing organism that inhabited shallow burrows at or near the sediment-water interface.

Over the past two decades, 16 additional fouled specimens of Flexicalymene spp. have been found in Upper Ordovician rocks of the Ohio-Indiana-Kentucky area. These specimens form the basis of this study and provide important new information that supports some of Brandt's (1996) conclusions, but they also suggest some alternative explanations. Our objectives are to: 1) describe the paleoenvironmental setting and lateral distribution of selected Upper Ordovician shale beds of the Ohio-Indiana-Kentucky area containing encrusted Flexicalymene spp. trilobites; 2) quantitatively describe the community composition, prevalence, abundance, and spatial distribution of epizoozoans and episkeletozoans on Flexicalymene spp.; 3) document preserved epizoozoan biotic interactions; 4) document preferred larval settling and alignment of epizoozoans; 5) discuss the feeding habits of epizoozoans and the host *Flexicalymene* spp.; and 6) describe the commensal relationships between these organisms.

MATERIALS AND METHODS

Nearly 15,000 articulated, relatively undeformed *Flexicalymene* spp. specimens were examined for epizoozoans and episkeletozoans from nine localities in the Cincinnati arch region of southwestern Ohio, southeastern Indiana, and northern Kentucky (Fig. 1). Most specimens probably belong to *F. meeki* (Foerste, 1910) or *F. retrorsa retrorsa* (Foerste, 1910) (see Hunda and Hughes, 2007). However, species-level identification was not attempted on non-figured specimens because numerous people, who were not all trilobite taxonomists, examined 1,000s of specimens for epizoans. Thus, the non-figured material is left under open nomenclature (i.e., *Flexicalymene* spp.) and from here on will be referred to simply as *Flexicalymene*.

The largest collection came from Mt. Orab, Ohio, where \sim 14,500 specimens were excavated during extensive quarrying of the trilobite shale bed (Ferree, 1994; Hunda et al., 2006). These specimens were examined for episkeletozoans with only the naked eye. Surface exposures at the remaining eight locations yielded 458 more specimens. These specimens were more carefully examined under a binocular microscope. All specimens were collected from six stratigraphic intervals, referred to as trilobite shale beds, within the Upper Ordovician Katian Stage (Maysvillan and Richmondian Stages of the historic North American Cincinnatian Series succession [Bergström et al., 2009]; Fig. 2). Prevalence of fouling was calculated as the percentage of fouled specimens at each location (Table 1). We also examined a smaller collection of well preserved specimens from the Treptoceras duseri shale exposed at Bevis, Ohio. In these specimens, width of the 1p preoccipital lobe was measured to the nearest 1 mm as a proxy for host size (Fig. 3).

We classified each fouled *Flexicalymene* as a carcass or exuvium using the criteria of Brandt (1993). A *Flexicalymene* carcass is characterized by intact librigenae and hypostome, and by thoracic segments that are not telescoped, displaced, or unevenly spaced. All 13 thoracic segments should be evident. Complete spheroidal enrollment is considered conclusive evidence of a carcass. *Flexicalymene* exuviae meet one or more of the following conditions: missing sclerites such as librigenae, hypostome, or other cephalic elements; incomplete enrollment or sharply bent cephala; displaced, disarticulated,



FIGURE 1—Locations of studied *Flexicalymene* Shirley, 1936 trilobites and outcrop pattern of trilobite shale beds from Upper Ordovician rocks of southwestern Ohio, southeastern Indiana, and northern Kentucky. The lateral distribution of the Madison trilobite shale bed is unknown. The Moranburg trilobite shale bed has been traced for 0.5 km.

telescoped, or unevenly spaced thoracic segments; or rotated pygidia or cephala.

For the fouled specimens, the degree of specimen compaction and its attitude (i.e., enrolled versus outstretched) were recorded. Width of the 1p preoccipital lobe and the dorsal surface area of the exoskeleton were measured on each fouled *Flexicalymene*. Epizoozoans and episkeletozoans were identified, counted, and examined for biotic interactions and preferred orientation. The surface area of each colonial or solitary episkeletozoan was measured. All 1p preoccipital lobe width measurements were made to the nearest 0.01 mm, and all surface area measurements were done with a digital image analysis system with measurement errors of less than 3%.

All fouled trilobite specimens of *Flexicalymene* are reposited in the Orton Geological Museum, The Ohio State University, Columbus, Ohio (OSU). They have been assigned catalog numbers OSU 53684 through OSU 53699 inclusive.

PALEOENVIRONMENTAL SETTING

The trilobite *Flexicalymene* ranges throughout the approximately 230 m of thin, interbedded limestone and shale beds characterizing the Cincinnatian succession (Caster et al., 1961;

Holland and Patzkowsky, 2007; Hunda and Hughes, 2007; Meyer and Davis, 2009). Flexicalymene and other trilobites commonly occur as disarticulated sclerites and average 9% of the bioclast fraction in Cincinnatian limestones (Martin, 1975). Trilobite bioclasts, ranging from 10 to 14% of fossil bioclasts, are most abundant in the deeper water, offshore environments of the Kope and Waynesville Formations and the intermediate water depths of the transitional, shoreface environments of the Fairview Formation, Corryville Member of the Grant Lake Formation, Corryville Member of the Grant Lake Limestone, and Liberty Formation. Trilobite bioclasts represent only 5 to 7% of fossil bioclasts in the shallower water, upper shoreface environments of the Bellevue and Mount Auburn Members of the Grant Lake Formation, the Bellevue and Straight Creek Members of the Grant Lake Limestone, the Arnheim Formation, and the Whitewater Formation, and represent only 1% of fossil bioclasts present in the lagoonal environments of the Saluda Formation (Martin, 1975).

Articulated *Flexicalymene* carcasses and exuviae occur throughout the Cincinnatian shales and limestones. However, the frequency of articulated *Flexicalymene* specimens is generally low in most stratigraphic intervals. So called



FIGURE 2—North American stratigraphic nomenclature of Upper Ordovician rocks of southwestern Ohio, southeastern Indiana, and northern Kentucky illustrating the positions of the *Flexicalymene* Shirley, 1936 trilobite shale beds and the number of fouled specimens collected.

"trilobite shale beds" are an exception to the general pattern of occurrence because these beds are characterized by abundant articulated trilobite carcasses and exuviae, complete crinoid crowns, butterflied bivalves, and complete articulated ostracodes (Brandt, 1985; Frey, 1987a, 1987b; Schumacher and Shrake, 1997; Hughes and Cooper, 1999; Hunda et al., 2006). These shales are thick-bedded, laminated claystones to lime mudstones with occasional, thin, fining upward laminae and beds of skeletal debris. Most articulated fossils and disarticulated fossil debris occur in clusters or patches separated by largely unfossiliferous claystone or mudstone. Trilobite shale beds occur as localized beds having limited geographic distribution (e.g., Hughes and Cooper, 1999), or beds that are regionally persistent and laterally traceable for tens of kilometers (e.g., Frey, 1987a, 1987b; Schumacher and Shrake, 1997). Brandt (1985), Frey (1987a, 1987b), Schumacher and Shrake (1997), Hughes and Cooper (1999), Babcock (2005), and Hunda et al. (2006) viewed storm related sedimentation as responsible for smothering benthic communities that are now preserved in trilobite shale beds. Rapid burial apparently inhibited scavenging and skeletal disarticulation, resulting in excellent preservation of trilobites and other fossils.

SAMPLE LOCATIONS AND STRATIGRAPHY

Flexicalymene specimens examined in this study occur in six discrete trilobite shale beds exposed at nine locations (Fig. 1). The stratigraphic interval containing the trilobite shale beds ranges from the basal Corryville Member of the Grant Lake

TABLE 1-Prevalence of episkeletozoans on Flexicalymene from Cincinnatian trilobite shale beds.

Number of <i>Flexicalyment</i> specimens examined	e Number of fouled <i>Flexicalymene</i> specimens	Prevalence (%)	Trilobite shale bed	Location	Reference
$2,030 \\ \sim 14,500 \\ 383 \\ 37 \\ 12 \\ 11 \\ 6 \\ 4 \\ 3 \\ 2 \\ \sim 14,958$	8 3 5 2 1 1 1 1 1 1 1 1 1 6	$\begin{array}{c} 0.39 \\ 0.02 \\ 1.3 \\ 5.4 \\ 8.3 \\ 9.1 \\ 17 \\ 25 \\ 33 \\ 50 \\ 0.11 \end{array}$	Multiple beds Mt. Orab Treptoceras duseri Madison Western Hills Western Hills Dent Moranburg Treptoceras duseri	Cincinnati, OH region Georgetown, OH Bevis, OH Caesar Creek, OH Madison, IN Queenswood Road, OH Quail Hill Road, OH Dent, OH Maysville, KY Weisburg, IN Total this study	Brandt (1996) This study This study This study This study This study This study This study This study This study This study



FIGURE 3—Size-frequency histogram of the *Flexicalymene* Shirley, 1936 population collected from the *Treptoceras duseri* shale exposed at Bevis, Ohio, compared to the size-frequency of the 16 fouled *Flexicalymene* specimens.

Limestone to the middle Waynesville Formation of Schumacher et al. (1991). In terms of historic North American stratigraphic nomenclature, the trilobite shale beds range from the middle Maysvillian Stage to the lower Richmondian Stage of the Cincinnatian (C3 to C5 third order sequences of Holland and Patzkowsky, 1996; Fig. 2). This interval correlates to the Katian Stage of the Upper Ordovician Series as now used globally (Bergström et al., 2009).

The Moranburg trilobite shale bed is exposed in two roadcuts along U.S. Route 62, 3.2–3.7 km northeast of the junction of Kentucky Routes 9 and 10 near Maysville, Kentucky (Fig. 1). An 11 cm thick claystone, containing scattered pockets of well preserved *Glyptocrinus* sp. Hall, 1847, occurs in the basal Corryville Member, about 1 m above the top of the Bellevue Member of the Grant Lake Limestone (Fig. 2). The total lateral extent of this bed is unknown but it is easily traced for 0.5 km along the high ridge traversed by U.S. Route 62.

The Western Hills trilobite shale bed crops out in natural stream exposures and construction site excavations in the Cincinnati, Ohio, area (Fig. 1). This shale occurs in the upper part of the Corryville Member of the Grant Lake Formation between 2 and 3 m below the Corryville-Mount Auburn Member contact (Fig. 2). This 1 m claystone is traceable for at least 20 km from the Quail Hill and Queenswood locations northward to Bevis, Ohio.

The Dent trilobite shale bed (*Glyptocrinus* sp. beds of Caster, 1961) is a 1 m thick, medium- to thick-bedded mudstone interbedded with thin limestones in the basal Arnheim Formation (Fig. 1). The base of this shale occurs about 1.5 m above the top of the Mount Auburn Member of the Grant Lake Formation (Fig. 2). This shale has a lateral distribution similar to the Western Hills trilobite shale bed (Fig. 1).

The informal Mt. Orab shale bed of Hunda et al. (2006) has been quarried from extensive excavations adjacent to Flat Run north of Georgetown, Ohio (Fig. 1). This trilobite shale bed consists of 0.46 m of gray lime mudstone interbedded with

nodular- and wavy-bedded calcisiltite and limestone. The Arnheim Formation index fossils, Retrorsirostra carleyi (Hall, 1860) and Leptaena richmondensis Foerste, 1909b, are present in the overlying nodular limestone and shale beds (Ferree, 1994; Hunda et al., 2006). Geologic mapping of Brown and Highland counties places the base of the Mt. Orab shale bed approximately 10 m above the Arnheim-Grant Lake Limestone formational contact (e.g., Schumacher et al., 2000). Weir et al. (1980) described a similar shale bed containing abundant, articulated trilobites (Flexicalymene and Isotelus sp. Dekay, 1824) overlain by a thin interval of nodular-bedded limestones and shales containing R. carlevi and L. richmondensis from stream exposures of the Arnheim Formation at its type section in Straight Creek near Arnheim, Ohio (Fig. 1). The Mt. Orab shale bed has a lateral distribution of at least 15 km from Flat Run Creek to Straight Creek.

The Madison trilobite shale bed is exposed in roadcuts of U.S. Route 421 approximately 2 km northeast of the intersection of Indiana Route 56 and U.S. Route 421 near Madison, Indiana. The 1.5 m thick blue shale is located near the top of the Arnheim Formation (Hattin et. al., 1961), which is also correlated to the middle of the Dillsboro Formation (Brown and Lineback, 1966). The lateral distribution of this shale is unknown.

The *Treptoceras duseri* shale of southwestern Ohio (sensu Flower, 1946) and its lateral equivalent, the trilobite shale unit of southeastern Indiana (sensu Frey, 1987a, 1987b), is exposed at Bevis, Ohio, Caesar Creek, Ohio, and Weisburg, Indiana (Fig. 1). This unit consists of approximately 1.5 m of thickbedded claystone with thin-bedded limestone capped by the *Onniella meeki* Miller, 1875 zone consisting of 1.5 to 2.5 m of interbedded shale and limestone. Frey (1997) determined the base of the *T. duseri* shale to be 10 to 11 m above the Arnheim-Waynesville formational contact in southwestern Ohio (Fig. 2). The trilobite shale unit occurs in the upper Dillsboro Formation in southeastern Indiana. Frey (1997) traced the *T. duseri* shale, *O. meeki* zone, and trilobite shale unit for 135 km through southwestern Ohio and southeastern Indiana (Fig. 1).

RESULTS

Of the ~14,958 *Flexicalymene* specimens examined, 16 (0.1%) were fouled. Eleven (69%) of these were preserved enrolled or semi-enrolled, and five (31%) were outstretched. Thirteen (81%) of the fouled specimens are considered carcasses, and three (19%) are interpreted as nearly intact exuviae (Table 2). Fouled specimens of *Flexicalymene* have cephala ranging from 6.61 to 12.90 mm (mean = 9.50 mm) across the width of the 1p preocciptal lobe. This size range suggests that these specimens ranged from early holaspids to gerontic specimens well past their terminal molt. Of the 383 *Flexicalymene* specimens collected from the Bevis, Ohio, location, 313 were not deformed, and their 1p widths could accurately be measured. These ranged from 2 to 14 mm, and all are holaspids (Fig. 3).

The community of commensal episkeletozoans consisted of three trepostome bryozoan genera, three brachiopod species, one annelid species, and a tube-dwelling/boring organism. Nine of the *Flexicalymene* specimens were fouled by a total of 50 trepostome bryozoan colonies (Table 2) belonging to *Parvohallopora* sp. Singh, 1979 (40 colonies), *Amplexopora* sp. Ulrich, 1882 (6 colonies), and *Heterotrypa* sp. Nicholson, 1879 (4 colonies). More bryozoans occurred on carcasses than exuviae but the differences are not significant because of the small number of fouled exuviae (*t*-test, P = 0.167). By contrast, more brachiopods occurred on exuviae than

TABLE 2-Enrollment attitude, fossil type, and size information for the	e 16 fouled specimens of <i>Flexicalymene</i> and their episkeletozoans.
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<i>Flexicalymene</i> specimen number	Enrollment attitude	Fossil type	lp width (mm)	Projected or measured dorsal surface area of host carapace (mm ²)	Number of bryozoan colonies	Total surface area of all bryozoan colonies (mm ²)	Maximum bryozoan colony height (mm) above host carpace
OSU 53684	enrolled	carcass	9.10	491	10	82.47	4
OSU 53685	prone	carcass	9.93	607	0		
OSU 53686	semi-enrolled	exuvia	11.34	612	0		
OSU 53687	enrolled	carcass	11.50	621	3	109.61	$\frac{2}{7}$
OSU 53688	prone	carcass	7.50	405	1	158.42	7
OSU 53689	prone	carcass	8.35	460	0		
OSU 53690	prone	exuvia	6.61	309	0		
OSU 53691	enrolled	carcass	6.76	365	0		
OSU 53692	enrolled	carcass	6.64	359	0		
OSU 53693	enrolled	carcass	10.44	564	0		
OSU 53694	enrolled	carcass	7.50	405	2	19.48	1
OSU 53695	enrolled	carcass	10.95	591	1	47.47	2
OSU 53696	enrolled	carcass	9.90	535	1	7.54	1
OSU 53697	semi-enrolled	exuvia	11.11	600	1	18.93	1
OSU 53698	enrolled	carcass	12.90	697	1	23.07	1
OSU 53699	prone	carcass	11.50	621	30	55.35	2
Mean:	•		9.50	515	3.1	58.0	
Sum:				8242	50	522	
Minimum:			6.61	309	0	7.54	
Maximum:			12.90	697	30	158.42	
Standard deviation:			2.04	117	7.6	50.30	

carcasses but the differences are not significant (*t*-test, P = 0.422).

Two *Flexicalymene* exoskeletons were encrusted with a total of six specimens of the inarticulate brachiopods *Schizocrania filosa* Hall, 1847 and *Petrocrania scabiosa* (Hall, 1868; Table 2). One *Flexicalymene* was colonized by at least 124 individuals of the articulate brachiopod *Zygospira modesta* (Say in Hall, 1847). Five *Flexicalymene* specimens were encrusted with 22 calcareous *Cornulites corrugatus* (Hall, 1888) annelid tubes. More *C. corrugatus* occurred on carcasses than exuviae but the differences are not significant because of the small number of fouled exuviae (*t*-test, P = 0.111).

On average there were more brachiopods (mean = 8.1 per fouled host) than bryozoans (mean = 3.1) or worm tubes (mean = 1.3). These numbers are skewed by one exuvium that was fouled by 124 brachiopods. Excluding all the exuviae (i.e., using the remaining 13 carcasses), there were more bryozoans per host (mean = 3.8) than worm tubes (mean = 1.5) or brachiopods (mean = 0.4). On average bryozoans covered more of their hosts' dorsal surface area (mean = 38.7 mm^2) than worm tubes (mean = 1.24 mm^2) or brachiopods (mean = 0.92 mm^2). The cumulative percent of the host carcass' dorsal surface area that was covered by all epizoozoans ranged from 0.1% to 39.1% (mean = 8%). Overall, the amount of surface area of hosts that were covered by epizoozoans was positively and significantly correlated with the 1p preoccipital lobe width $(\mathbf{R}^2 = 0.014, P = 0.035)$. This suggests that larger fouled trilobites provided more area for the epizoozoans to grow.

Sizes (as measured by both 1p preoccipital lobe width and dorsal surface area) of fouled carcasses and exuviae are not significantly different (*t*-tests, P > 0.05). Therefore any differences in the prevalence of fouling of carcasses and exuviae are not simply a function of different size targets for larval settlement.

We excluded exuviae from the following analyses because the shed molts may represent older substrates that were fouled by larvae at various times. The carcasses were more likely to be fouled while the hosts were alive. The carcasses of fouled hosts were significantly larger (mean 1p width = 10.25 mm) than the non-fouled carcasses (mean = 6.75 mm; *t*-test, P =0.043), indicating that the larger trilobites were more likely to be fouled by settling larvae. This is expected because larger trilobites molted less frequently than smaller (i.e., younger) trilobites (Hunt, 1967; Chatterton et al., 1994; Clarkson et al., 2003). Also, larger trilobites provided larger targets for settling larvae.

The prevalence of epizoozoans and episkeletozoans fouling *Flexicalymene* varied between collection localities from 0.02% to 50% (Table 1). We suggest that encrustation frequencies of less than 2% reflect the normal rate of fouling for *Flexicalymene* in the Late Ordovician. Frequencies greater than 2% are most likely due to sampling bias resulting from small samples.

The Heterotypa sp. and Amplexopora sp. epizoozoans fouling Flexicalymene retrorsa retrorsa specimens collected from two Ohio localities (Mt. Orab and Bevis) contain numerous cylindrical tubes or borings incorporated in the bryozoan colonies (Fig. 4). The tubes/borings are oriented perpendicularly to the surface of the colony, and range in diameter from 0.2 to 0.8 mm. The bryozoan zooids parallel the tube/boring boundaries. Some tube/boring openings are surrounded by zooids forming an elevated ring around each tube/boring. The elevated rims suggest the bryozoan was still alive when the tube/boring formed. If so, this is an example of a bioclaustration (Tapanila, 2005). The borings without the elevated rims are similar to the trace fossil Sanctum sp. Erickson and Bouchard, 2003 (Wyse Jackson and Key, 2007). Identification of the exact trace fossil is not certain as thin sectioning of the bryozoan colony was not possible. If it is Sanctum sp., this is the first reported fossil occurrence of a sessile organism (Heterotrypa sp.) encrusting a live motile host (Flexicalymene) that itself is bored by a presumed filter feeding polychaete, the probable tracemaker of Sanctum sp. (Wyse Jackson and Key, 2007).

The Mt. Orab *Heterotrypa* sp. colony (Fig. 4.1, 4.2) incorporated 25 tubes/borings concentrated on the elevated portion of the colony encrusting the cranidium of one *Flexicalymene retrorsa retrorsa* specimen and the upper part of the single branch of the bryozoan colony. The *Amplexopora* sp. colony from Bevis (Fig. 4.3) incorporated five tubes/ borings along the anterior margin of the doublure. The tubes/ borings point forward, not upward, as observed in the Mt. Orab *Heterotrypa* sp. colony.

Epizoozooans and episkeletozoans are not randomly distributed on the 16 fouled *Flexicalymene* specimens, but

TABLE 2—Extended.

Bryozoan genus	Number of brachiopods	Total surface area of all brachiopods (mm ²)	Number of <i>Cornulites</i>	Total surface area of all <i>Cornulites</i> (mm ²)	Total number of episkeletozoans	Total surface area of all episkeletozoans (mm ²)	% of dorsal surface area of host carapace covered by episkeletozoans
Parvohallopora	0		0		10	82.47	16.8
*	0		1	2.46	1	2.46	0.4
	1	29.65	0		1	29.65	4.8
Heterotrypa	0		0		3	109.61	17.7
Heterotrypa	0		0		1	158.42	39.1
	0		11	9.55	11	9.55	2.1
	124	34.23	0		124	34.23	11.1
	5	11.93	0		5	11.93	3.3
	0		1	0.26	1	0.26	0.1
	0		8	0.72	8	0.72	0.1
Amplexopora	0		0		2	19.48	4.8
Amplexopora	0		0		1	47.47	8.0
Amplexopora?	0		0		1	7.54	1.4
Amplexopora	0		0		1	18.93	3.2
Amplexopora	0		1	1.94	2	25.01	3.6
Parvohallopora	0		0		30	55.35	8.9
*	8.1	25.27	1.4	2.99	12.6	38.32	7.8
	130	75.81	22	14.9	202	613	
	0	11.93	0	0.26	1	0.26	0.1
	124	34.23	11	9.55	124	158.42	39.1
	30.9	11.8	3.2	3.78	30.6	44.3	10.0

preferentially encrust the elevated regions of the exoskeleton. On the 13 carcasses, 55 (65%) of the epizoozoans encrusted the glabella, fixigenae adjacent to the glabellar furrow, occipital ring, eye ridges, axial rings, fulcra of the pleurae, and axial rings of the pygidum (Table 3; Figs. 5, 6). Sixteen (19%) epizoozoans were attached to the anterior margin of the cephalon and doublure. The remaining 13 (15%) epizoozoans were preserved fouling the librigena, pleural field, distal portions of individual pleura, or the axial furrows (Table 3; Fig. 7).

The three inferred exuviae of *Flexicalymene* display a pattern of episkeletozoan fouling similar to that on the carcasses. Ninety one (72%) episkeletozoans encrusted the elevated regions of the glabella, fixigenae adjacent to the lateral glabellar furrows, axial rings, and fulcra of the pleurae (Table 3; Figs. 5, 6). Thirty five (28%) episkeletozoans were preserved attached to the lower regions of the librigena, pleural fields, or within the axial furrows (Table 3; Figs. 8, 9).

Parvohallopora sp. and *C. corrugatus* display preferred alignment of individual colony branches or individual tube apertures. One *Flexicalymene retrorsa retrorsa* specimen (Fig. 6) has five parallel, aligned *Parvohallopora* sp. colonies. The colonies are centered on the eye ridges, the glabella, and either side of the doublure. The colony branches did not grow vertically from the cephalon, but instead grew more or less horizontally beyond the anterior margin of the cephalon (Fig. 6.1, 6.2).

One *Flexicalymene meeki* specimen (Fig. 5) contains 30 *Parvohallopora* sp. colonies of which 28 (90%) occur on the elevated regions of the glabella, eye ridges, pleural fulcra and fields, and axial rings immediately adjacent to the glabellar, axial, and pleural furrows. On the pleurae, each colony grew along the axis of a pleuron but did not extend to the neighboring pleuron. Fourteen (45%) of these colonies contain 1–3 mm long branches that grew laterally at angles ranging from 30° to 45° above horizontal. Branches of all 14 colonies are aligned parallel to the axis of each pleuron, each occipital or axial ring, or the posterior margin of the cephalon. They also extend laterally into the area above the glabellar, axial, and pleural furrows or from the pleural fulcra toward a lateral margin of the specimen.

The tubes of *C. corrugatus* display a variety of orientations at their initial attachment site on the cephalons of the five fouled *Flexicalymene* specimens. Fourteen (70%) of 20 specimens bend so that the aperture opens posteriorly and parallel to the sagittal axis (Fig. 7). Five (25%) specimens are oriented with the aperture opening anteriorly, and one (5%) is oriented with the aperture oriented perpendicularly to the sagittal axis.

DISCUSSION AND CONCLUSIONS

The diversity of marine hard substrate encrusters on Flexicalymene is somewhat less than the hard substrate community encrusting sessile epifaunal organisms of the Cincinnatian sea floor. Previous studies of Cincinnatian fossils have documented that sessile epifaunal organisms were fouled by articulate and inarticulate brachiopods, trepostome bryozoans, cornulitids, rugose and tabulate corals, crinoids, edrioasteroids, and stromatoporoids (Richards, 1972; Alexander and Scharpf, 1990; Meyer, 1990). Motile Cincinnatian organisms interpreted as having been fouled by encrusting and ramose bryozoans and cornulitids while living include cephalopods, gastropods, monoplacophorans, and trilobites (Morris and Rollins, 1971; Baird et al., 1989; Morris and Felton, 1993; Brandt, 1996). Our investigation adds inarticulate and articulate brachiopods and an undetermined tube-dwelling/boring organism, probably Sanctum sp., to the hard substrate community living on Late Ordovician Flexicalymene.

Why are epizoozoans so rare on trilobites in general and on *Flexicalymene* in particular? Brandt (1996), who previously reported a 0.4% prevalence of encrustation from a large museum collection of *Flexicalymene* specimens, concluded that the low prevalence of fouling is a function of opportunistic larval settlement, preservational bias resulting from the loss of nonbiomineralized epizoozoans, and a semi-infaunal, shallow burrowing lifestyle of *Flexicalymene*. Studies of antifouling mechanisms in modern crabs have shown that burrowing plays the largest role in antifouling (e.g., Becker and Wahl, 1996). Shallow burrowing by *Flexicalymene* no doubt limited the amount of area of the dorsal exoskeleton available for larval settling, in part because of the scouring action of sediment (Smith and Nelson, 2003).



FIGURE 4—1–3, Flexicalymene retrorsa retrorsa (Foerste, 1910) fouled by Heterotrypa sp. Nicholson, 1879 and Amplexopora sp. Ulrich, 1882 bryozoan colonies containing tube-dwelling/boring nonbiomineralized organisms including Sanctum sp. Erickson and Bouchard, 2003; 1, F. retrorsa retrorsa, OSU 53688 from Mt. Orab trilobite shale bed exposed near Georgetown, Ohio, fouled with Heterotrypa sp. bryozoan colony covering ~80% of the cephalon and containing 25 vertical tubes/borings, dorsal view, 1p width: 7.50 mm; 2, OSU 53688, tube-dwelling/boring organisms are restricted to the elevated portions of the colony, lateral view, 1p width: 7.50 mm; 3, F. retrorsa retrorsa, OSU 53695 from the Treptoceras duseri shale exposed at Bevis, Ohio, encrusted along the anterior margin by an Amplexopora sp. bryozoan colony and five, anteriorly directed, horizontally oriented tubes/borings, anterior view, 1p width: 10.95 mm.

Conceivably, the low prevalence of fouling of *Flexicalymene* by bryozoans, brachiopods, and annelid worms reflects a larval preference for, or a greater opportunity to settle on, other substrates. Larvae are known to respond to various aspects of substrates including mechanical properties (e.g., Gray et al., 2002) and/or the presence of microbial biofilms (e.g., Kitamura and Hirayama, 1987). The larval settlement on these trilobites may have been purely accidental and phoretic (i.e., hitchhiking). A low intensity of fouling has been found with bryozoans on extant sea snakes (0.8%: Key et al., 1995),

			Topogra	graphically elevated regions of carapace	evated regi	ons of caral	ace			Topographi	Topographically depressed regions of carapace	sed regions	of carapace	
	Flexicalymene		Cephal	lon		The	Thorax	Pygidium		Cephalon		Thorax	ax.	Pygidium
Episkeletozoan	specimen	Glabella	Fixigena	Occipital ring	Eye ridges	Axial rings	Pleura fulcra	Axial rings	Anterior margin	Librigena	Doublure	Axial furrows	Pleura	Pleura field
Amplexopora, Heterotrypa, and Parvohallopora	OSU 53684 OSU 53687 OSU 53687 OSU 53694 OSU 53694		$\begin{array}{c} 1\\ 0\\ 1^{a}\end{array}$	1 1 1 0 1 1 0	0000	0000	0000	0000	-100 ⁴	0 1 ^a 0 0	0000	0000	0000	~000
	OSU 53695 OSU 53696 OSU 53697 OSU 53697 OSU 53698	00	0000	0000	0000	0000	0000	0000	1 ^b 001 ^b	0000	0000	0000	0000	0000
Cornulites corrugatus	OSU 53699 Sum: OSU 53685 OSU 53689 OSU 53693 OSU 53693	$10^{-3}, 10^{-1}, 10^{-3}, 10^{-1}, 1$	040000	- x 0 - q	- v 0 % 0 0 0	∞∞०००००	000-000	~~~~~~	1 6 0 1 ^c 1 ^c	000-000	000-00-	000000	00000	000000
Schizocrania filiosa and Petrocrania scabiosa	Sum: Sum: OSU 53686 OSU 53691	0610-	0000	0-000	00000	00000	0-000		00000	01000	-000	000m		
Zygospira modesta	Sum: OSU 53690 Grand total:	$13 \\ 33$	11 5 4	004	00%	0 35 35	042	00m	0 0 11	0 7 11	000	25 28	D (n 4	006
^a colony covers most of cephalon. ^b colony extends onto doublure	ephalon. ^b color	iy extends o	onto doublur	ి.	aperture facing anterior.	p	ture facing	aperture facing posterior. ^e aperture facing perpendicular to host's sagittal axis.	aperture fac	ing perpend	icular to hos	st's sagittal a	axis.	



FIGURE 5—1–3, Flexicalymene meeki (Foerste, 1910), OSU 53699 from the Moranburg trilobite shale bed exposed at Maysville, Kentucky fouled with juvenile Parvohallopora sp. Singh, 1979 bryozoan colonies; *1*, Parvohallopora sp. bryozoan colonies preferentially fouling the cranidium, eye ridges, axial rings, and pleural fulcra adjacent to the axial furrows; Lateral deformation and attitude of the cephalon obscures some of the Parvohallopora sp. bryozoan colonies on the right librigena and pleura, dorsal view, 1p width: 11.50 mm; 2, Locations of 28 of 30 Parvohallopora sp. bryozoan colonies on the elevated portions of the exoskeleton of OSU 53699 mapped onto a dorsal view of an unfouled specimen of *Flexicalymene* sp. modified from Babcock (2005, fig. 8–1.10); 3, Locations of he cephalon of OSU 53699 mapped onto an anterior view of an unfouled specimen of *Flexicalymene* sp. modified from Babcock (2005, fig. 8–1.8).

but most bryozoan fouling rates are higher on extant arthropods such as blue crabs (16%: Key et al., 1999), isopods (42%: Key and Barnes, 1999), and horseshoe crabs (57–77%: Key et al., 1996a, 1996b, 2000). Studies by Richards (1972), Alexander and Scharpf (1990), and Meyer (1990) show that in the Cincinnatian, encrusters colonized any available hard substratum whether pebbles or cobbles, motile or sessile organisms, or living or dead hosts. If so, larval settlement on *Flexicalymene* was probably greatly influenced by exposure of its exoskeleton above the sediment water interface and chemical signaling. As a motile, burrowing animal, exoskeletons of *Flexicalymene* may have been only intermittently available for larval settlement.

Is the low prevalence of fouling of *Flexicalymene* related to frequent molting? With a sample size of roughly 15,000 in all holaspid size ranges showing low rates of encrustation, this possibility can largely be ruled out.

There is a possibility that the low prevalence of fouling is an observational bias. Lescinsky (1996) noted that episkeletozoans are commonly overlooked because of small size or because of removal along with matrix adhering to fossil specimens during preparation. Waugh et al. (2004) stated that episkeletozoans can go unreported in taxonomic studies because the presence of episkeletozoans seems irrelevant to those studying the hosts. Observational bias may have



FIGURE 6—1–2, Preferred alignment of *Parvohallopora* sp. Singh, 1979 bryozoan colonies on *Flexicalymene retrorsa retrorsa* (Foerste, 1910), OSU 53684 from the *Treptoceras duseri* shale exposed at Caesar Creek, Ohio, 1p width: 9.10 mm; 1, alignment of *Parvohallopora* sp. bryozoan colonies originating from the glabella, occipital ring, eye ridges, anterior margin and paralleling the axial furrows, dorsal view; 2, anterior growth of *Parvohallopora* sp. bryozoan branches extending beyond anterior margin and parallel one another, lateral view.

contributed to the apparent low prevalence of fouling among *Flexicalymene* specimens collected from Mt. Orab, Ohio, as these specimens were inspected only with the naked eye and not thoroughly examined under a binocular microscope. Some tiny episkeletozoans were undoubtedly overlooked.

Preservational bias is another possible cause of the apparent low prevalence of fouling. Some episkeletozoans were almost certainly lost from *Flexicalymene* specimens as they weathered from the soft Cincinnatian shales. Further taphonomic bias could result from the degradation of *Flexicalymene* epicuticle, something that has been observed in fossil decapods (Waugh et al., 2004). Dalingwater et al. (1993) argued that the thin, outermost layer of trilobite exoskeleton was an epicuticle similar to that of decapods. Low prevalence of fouling observed in *Flexicalymene* may reflect the loss of the epicuticle along with its episkeletozoans before fossilization.





FIGURE 7—1–3, Cornulites corrugatus (Hall, 1888) annelid tubes encrusting the cephalon of *Flexicalymene retrorsa retrorsa* (Foerste, 1910). 1, A composite sketch from OSU 53685, OSU 53692, OSU 53693, and OSU 53698 from the Madison trilobite shale bed exposed at Madison, Indiana, and the *Treptoceras duseri* shale exposed at Caesar Creek, Ohio, illustrating a posterior preferred orientation of the apertures of 14 specimens of *C. corrugatus* mapped onto a dorsal and anterior view of an unfouled cephalon of *Flexicalymene* sp. modified from Harrington et al. (1959, fig. 354, 1a and 1c); 2, five smaller *C. corrugatus* with four displaying posteriorly oriented apertures and the fifth an anterior orientation, fouling the anterior margin of an enrolled *F. retrorsa retrorsa*, OSU 53695, dorsal view, 1p width: 9.93 mm.

Epibionts can serve to camouflage animals. Vance (1978) reported predation rates by starfish on clams were reduced when the clams were covered by bryozoans. It could be speculated that epizoozoans camouflaged *Flexicalymene* individuals from their predators, or alternatively from their prey. The low percentage of the hosts' dorsal surface area covered by epizoozoans in this study (mean = 8%) and the low incidence of epizoozoan encrustation, however, suggests this benefit did not accrue to *Flexicalymene*. Another more

speculative idea is that *Flexicalymene* harbored microbial symbionts that tended to discourage larval settlement. This is essentially untestable from fossil material however.

Successful larval settlement on Cincinnatian *Flexicalymene* shows a clear preferred pattern. Brandt (1996) observed that epizoozoans and episkeletozoans consistently fouled the elevated central regions of the dorsal exoskeleton and the anterior cephalic margin. Our results are consistent with those of Brandt (1996). Epizoozoans and episkeletozoans on



FIGURE 8—1–2, Schizocrania filosa Hall, 1847 brachiopods encrusting *Flexicalymene* sp. (Foerste, 1910). 1, A composite sketch from OSU 53686 and OSU 53691 from the *Treptoceras duseri* shale and the Western Hills trilobite shale bed illustrating six *S. filosa* and one *Cornulites corrugatus* (Hall, 1888) encrusting *Flexicalymene* sp. mapped onto a dorsal view of an unfouled specimen of *Flexicalymene* sp. modified from Babcock (2005, fig. 8-1.10), apices of each *S. filosa* specimen are oriented within the axial furrows of *Flexicalymene* sp. and the anterior margin of each valve aligns with the elevated portions of the exoskeleton (i.e., glabella, pleural fulcra, and fixigenae); 2, anterior margin of *S. filosa* oriented along the preglabellar furrows and the central area of the glabella of a semi-enrolled *F. retrorsa retrorsa*, OSU 53686 from the *T. duseri* shale exposed at Caesar Creek, Ohio, dorsal view, 1p width: 11.34 mm.

Flexicalymene preferred the elevated portions of the exoskeleton or the anterior cephalic margin. On *Flexicalymene* carcasses, 81% of the epizoozoans were preserved attached either to the elevated regions of the glabella, fixigenae, occipital rings, eye ridges, axial rings, and fulcra of individual pleura, or the anterior margin of the librigena, fixigenae, and doublure. On *Flexicalymene* exuviae, 77% of episkeletozoans were preserved attached either to the elevated areas of the glabella, fixigenae, axial rings, pleura fulcra, or the anterior regions of the librigena. The fouling pattern is similar on both carcasses and exuviae of *Flexicalymene*, suggesting that the exuviae were not subject to extensive additional colonization by episkeletozoans prior to burial and preservation.

Did epizoozoans fouling *Flexicalymene* preferentially align or colonize preferred areas of the exoskeleton to take advantage of currents created by locomotion or other life activities of the host? Previous studies of Morris and Rollins (1971), Baird et al. (1989), Morris and Felton (1993), and Galle and Parsley (1995) concluded that aligned epizoozoans preserved on motile cephalopods, tergomyans, gastropods, and hyolithids displayed preferential orientation to take advantage of the movement or feeding currents generated by the hosts.



FIGURE 9—1–2, Zygospira modesta (Say in Hall, 1847) brachiopods fouling the dorsal exoskeleton of a *Flexicalymene meeki* (Foerste, 1910) exuvium, OSU 53690 from the Western Hills trilobite shale bed exposed at the Queenswood Road location, Ohio, dorsal view, 1p width: 6.61 mm; 1, Z. modesta juveniles preserved in life position with pedicle openings adjacent to the *F. meeki* exoskeleton and concentrated along the elevated axial rings and pleura fulcrums; 2, cluster of adult *Z. modesta* in life position fouling the anterior margin and adjacent glabellar furrow.

Flexicalymene classically has been interpreted as having a benthic, semi-infaunal suspension feeder habit. This trilobite is presumed to have excavated shallow concave burrows that were used as filter chambers under the dorsal exoskeleton (Osgood, 1970; Seilacher, 1985; Brandt, 1996). If true, the epizoozoans on Flexicalymene were possibly oriented to take advantage of feeding currents generated by the trilobite (Brandt, 1996). Examination of Rusophycus pudicum Hall, 1852 trace fossils, which were constructed by Cincinnatian Flexicalymene, however, has revealed that the trilobiteconstructed traces often truncate traces left by infaunal worms. This suggests the *Flexicalymene* was an active carnivore (English and Babcock, 2007), at least in part. An interpretation of a carnivorous life habit, including hunting of infauna, is consistent with phylogenetic evidence, which suggests an affinity of the trilobites among the largely carnivorous arachnomorph arthropods (Fortey and Owens, 1999; Babcock, 2003; English and Babcock, 2007). Not all Rusophycus pudicum traces were necessarily hunting or suspension feeding traces, but the abundance of such traces in Cincinnatian strata indicates that Flexicalymene was an active nektobenthic swimmer and burrower. Preferential disposition of epizoozoans therefore likely reflects sites on the trilobite exoskeleton that were inflexible or weakly flexible. This same pattern is seen among epizoozoans on a variety of extant hosts (Key et al., 1995, 1996a, 1996b, 1997). Apertures and commissures of the encrusters may have aligned to either take advantage of currents generated while the host swam, or to help prevent fouling by sediment while the host dug into the substrate.

Nascimento and Alexander (1994), who conducted flume analyses on a model of Calymene celebra Raymond, 1916, discovered that at water velocities exceeding 8 to 10 cm/sec, turbulent flow was distinctly visible across the glabellar furrows and behind the occipital ring and pygidium. If we assume that similar turbulent water flows would have occurred across the exterior exoskeleton of Flexicalymene, then most epizoozoans would have preferred locations on the elevated areas adjacent to the zones of turbulent flow. Many animals minimize the energy required to feed by taking advantage of natural turbulent flow created by obstructions hindering water currents (Vogel, 1981). Thus, organisms would preferentially gather to take advantage of food suspended in the water column by eddies created downstream from the obstructions. It is possible that epizoozoans observed in our study preferentially settled in areas within or adjacent to turbulent water flows across the *Flexicalymene* exoskeleton to take advantage of food laden eddies. Regardless, the parameters of the flume study under represent the complexities of the life habit of Flexicalymene as inferred from trace fossils.

Were older adult specimens of *Flexicalymene* nearing or beyond the terminal molt stage subject to increased fouling as opposed to younger individuals? Kloc (1997) observed that epizoozoans fouling the trilobite *Dicranurus hamatus elegantus* were on specimens of different growth stages. Brandt (1996) reported that only large specimens of *Flexicalymene*, interpreted as older adults nearing the terminal molt stage, were fouled by epizoozoans and episkeletozoans. Our results indicate that fouling probably occurred throughout most of the holaspid period of *Flexicalymene* because we observed a range from 6 to 13 mm for the width of the preoccipital lobes of the fouled hosts. Brandt (1996) reported the preoccipital lobe widths to range from 10 to 13 mm among the eight specimens of *Flexicalymene* that she measured. Together, the results of Brandt (1996) and those reported here indicate that larger, although not necessarily gerontic, trilobites are more likely to be fouled by settling larvae. We interpret this as partly a reflection of the fact that larger trilobites provide larger targets for settling larvae and partly because trilobite size is proportional to molting frequency. Larger, older trilobites probably molted less frequently than smaller, younger trilobites (Hunt, 1967; Chatterton et al., 1994; Clarkson et al., 2003).

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REFERENCES

- ALEXANDER, R. R. AND C. E. BRETT. 1990. Symposium on Paleozoic epibionts: Introduction. Historical Biology, 4:151–153.
- ALEXANDER, R. R. AND C. D. SCHARPF. 1990. Epizoans on Late Ordovician brachiopods from southeastern Indiana. Historical Biology, 4:179–202.
- BABCOCK, L. E. 2003. Trilobites in Paleozoic predator-prey systems, and their role in reorganization of early Paleozoic ecosystems, p. 55–92. *In* P. H. Kelley, M. Kowalewski, and T. A. Hansen (eds.), Predator-prey interactions in the fossil record. Kluwer Academic/Plenum, New York.
- BABCOCK, L. E. 2005. Phylum Arthropoda, class Trilobita, p. 90–113. In R. M. Feldmann, and M. Hackathorn (eds.), Fossils of Ohio. Ohio Division of Geological Survey Bulletin, 70.
- BAIRD, G. C., C. E. BRETT, AND R. C. FREY. 1989. "Hitchhiking" epizoans on orthoconic cephalopods: Preliminary review of the evidence and its implications. Senckenbergiana Lethaea, 69:439–465.
- BECKER, K. AND M. WAHL. 1996. Behavior patterns as natural antifouling mechanisms of tropical marine crabs. Journal of Experimental Marine Biology and Ecology, 203:245–258.
- BERGSTRÖM, S. M, S. C. FINNEY, X. CHEN, D. GOLDMAN, AND S. A. LESLIE. 2006. Three new Ordovician global stage names. Lethaia, 39:287–288.
- BERGSTRÖM, S. M., X. CHEN, J. C. GUTIÉRREZ-MARCO, AND A. DRONOV. 2009. The new chronostratigraphic classification of the Ordovician System and its relations to major regional series and stages and to δ^{13} C chemostratigraphy. Lethaia, 42:97–107.
- BIGSBY, J. J. 1825. Description of a new species of trilobite. Journal of the Academy of Natural Sciences of Philadelphia, 4:365–368.
- BRANDT, D. S. 1985. Ichnologic, taphonomic, and sedimentologic clues to the deposition of Cincinnatian shales (Upper Ordovician), Ohio, U.S.A., p. 299–307. *In* H. A. Curran (ed.), Biogenic Structures: Their use in interpreting depositional environments. SEPM Special Publication 35.
- BRANDT, D. S. 1993. Ecdysis in *Flexicalymene meeki* (Trilobita). Journal of Paleontology, 67:999–1005.
- BRANDT, D. S. 1996. Epizoans on *Flexicalymene* (Trilobita) and implications for trilobite paleoecology. Journal of Paleontology, 70:442–449.
- BRETT, C. E. AND W. L. TAYLOR. 1997. The *Homocrinus* Beds: Silurian crinoid lagerstätten of western New York and southern Ontario, p. 181– 223. *In* C. E. Brett, and G. C. Baird (eds.), Paleontological events: Stratigraphic, ecological, and evolutionary implications. Columbia University Press, New York.
- BROWN JR., G. D. AND J. A. LINEBACK. 1966. Lithostratigraphy of the Cincinnatian Series (Upper Ordovician) in southeastern Indiana. American Association of Petroleum Geologists Bulletin, 50:1018–1023.
- BUDIL, P. AND R. SARIC. 1995. Cemented epibionts on the exoskeleton of the odontopleurid trilobite *Selenopeltis vultuosa tenyl* Šnajdr, 1984. Věstník Českého Geologického Ústavu, 70:29–31.
- CAMPBELL, K. S. W. 1977. Trilobites of the Haragan, Bois d'Arc and Frisco Formations (Early Devonian), Arbuckle Mountains region, Oklahoma. Bulletin Oklahoma Geological Survey, 123:1–227.
- CASTER, K. E. 1961. Field trip 7: Sunday all-day field excursion in the Cincinnati region, p. 255–257. In Guidebook for field trips, Cincinnati meeting. Geological Society of America.

- CASTER, K. E., E. A. DALVÉ, AND J. K. POPE. 1961. Elementary guide to the fossils and strata of the Ordovician in the vicinity of Cincinnati, Ohio. Cincinnati Museum of Natural History, 47 p.
- CHATTERTON, B. D. E., G. D. EDGECOMBE, S. E. SPEYER, A. S. HUNT, AND R. A. FORTEY. 1994. Ontogeny and relationships of Trinucleoidea (Trilobita). Journal of Paleontology, 68:523–540.
- CLARKSON, E. N. K. AND R. P. TRIPP. 1982. The Ordovician trilobite *Calyptaulax brongniartii* (Portland). Transactions of the Royal Society of Edinburgh (Earth Sciences), 72:287–294.
- CLARKSON, E. N. K., J. AHLGREN, AND C. M. TAYLOR. 2003. Structure, ontogeny, and moulting of the olenid trilobite *Ctenopyge (Eoctenopyhe)* angusta Westergard, 1922 from the Upper Cambrian of Vastergotland, Sweden. Palaeontology, 46:2–27.
- DALINGWATER, J. E., S. J. HUTCHINSON, H. MUTVEI, AND D. J. SIVETER. 1993. Cuticular ultrastructure of some Silurian calymenid trilobites from the Welsh Borderland and Gotland. Palaeontographica, Abteilung A (Palaeozoologie-Stratigraphie), 229:37–49.
- DEKAY, J. E. 1824. Observations on the structure of trilobites, and descriptions of an apparently new genus. With notes on the geology of Trenton Falls by J. Renwick. Annals of the Lyceum of Natural History of New York, 1:174–189.
- ENGLISH, A. M. AND L. E. BABCOCK. 2007. Feeding behaviour of two Ordovician trilobites inferred from trace fossils and nonbiomineralised anatomy, Ohio and Kentucky, USA. Memoirs of the Association of Australasian Palaeontologists, 34:537–544.
- ERICKSON, J. M. AND T. D. BOUCHARD. 2003. Description and interpretation of *Sanctum laurentiensis*, new ichnogenus and ichnospecies, a domichnium mined into late Ordovician (Cincinnatian) ramose bryozoan colonies. Journal of Paleontology, 77:1002–1010.
- FERREE, R. A. 1994. Taphonomy, paleoecology, and depositional environment of a trilobite lagerstätten, Mount Orab, Ohio. Unpublished M.S. thesis, University of Cincinnati, 98 p.
- FLOWER, R. H. 1946. Ordovician cephalopoda of the Cincinnati region. Pt. I. Bulletins of American Paleontology, 29:83–738.
- FOERSTE, A. F. 1909a. Fossils from the Silurian formations of Tennessee, Indiana, and Kentucky. Denison University Science Laboratories Bulletin, 14:61–107.
- FOERSTE, A. F. 1909b. Preliminary notes on Cincinnatian and Lexington fossils. Denison University Science Laboratories Bulletin, 14:289–334.
- FOERSTE, A. F. 1910. Preliminary notes on Cincinnatian and Lexington fossils of Ohio, Indiana, Kentucky, and Tennessee. Denison University Science Laboratories Bulletin, 16:17–100.
- FORTEY, R. A. AND R. M. OWENS. 1999. Feeding habits in trilobites. Palaeontology, 42:429–465.
- FREY, R. C. 1987a. The occurrence of pelecypods in Early Paleozoic epeiric-sea environments, Late Ordovician of the Cincinnati, Ohio area. Palaios, 2:3–23.
- FREY, R. C. 1987b. The paleoecology of a Late Ordovician shale unit from southwest Ohio and southeastern Indiana. Journal of Paleontology, 61:242–267.
- FREY, R. C. 1997. The utility of epiboles in the regional correlation of Paleozoic epeiric sea strata: an example from the Upper Ordovician of Ohio and Indiana, p. 335–368. *In C. E. Brett and G. C. Baird (eds.)*, Paleontological Events: Stratigraphic, Ecological, and Evolutionary Implications. Columbia University Press, New York.
- GALLE, A. AND R. L. PARSLEY. 2005. Epibiont relationships on hyolithids demonstrated by Ordovician trepostomes (Bryozoa) and Devonian tabulates (Anthozoa). Bulletin of Geosciences, 80:125–138.
- GILI, J. M., P. ABELLO, AND R. VILLANUEVA. 1993. Epibionts and intermoult duration in the crab *Bathynectes piperitus*. Marine Ecology Progress Series, 98:107–113.
- GRAY, N. L., W. C. BANTA, AND G. I. LOEB. 2002. Aquatic biofouling larvae respond to differences in the mechanical properties of the surface on which they settle. Biofouling, 18:269–273.
- HALL, J. 1847. Palaeontology of New York. Vol. 1. Containing Descriptions of the Organic Remains of the Lower Division of the New York System (Equivalent of the Lower Silurian Rocks of Europe). Natural History of New York, State of New York, Albany, 338 p.
- HALL, J. 1852. Palaeontology of New York. Vol. 2. State of New York, Albany, 362 p.
- HALL, J. 1860. Contributions to palaeontology, 1858 and 1859. Annual report of the Regents of the University, on the condition of the New York State Cabinet of Natural History, 13:53–125.
- HALL, J. 1868. New or little known species of fossils from rocks of the age of the Niagara Group. Annual report of the Regents of the University, on the condition of the New York State Cabinet of Natural History, 20:347–438.
- HALL, J. 1888. Palaeontology of New York. Vol. 7. Supplement containing descriptions and illustrations of Pteropoda, Cephalopoda

and Annelida. Geological Survey of the State of New York, C. van Benthuysen and Sons Albany, 42.

- HARRINGTON, H. J., G. HENNINGSMOEN, B. F. HOWELL, V. JAANUSSON, C. LOCHMAN-BALK, R. C. MOORE, C. POULSEN, F. RASETTI, E. RICHTER, R. RICHTER, H. SCHMIDT, K. SDZUY, W. STRUVE, L. STØRMER, C. J. STUBBLEFIELD, R. TRIPP, J. M. WELLER, AND H. B. WHITTINGTON. 1959. Arthropoda 1, Trilobitomorpha. Pt. O. In R. C. Moore (ed.), Treatise on Invertebrate Paleontology. Geological Society of America, New York, and University of Kansas Press, Lawrence.
- HATTIN, D. E., E. NOSOW, R. D. PERKINS, E. C. STUMM, M. C. MOUND, AND J. UTGAARD. 1961. Field trip 9: field excursion to the Falls of the Ohio, p. 295–350. *In* Guidebook for field trips, Cincinnati meeting. Geological Society of America.
- HOLLAND, S. M. AND M. E. PATZKOWSKY. 1996. Sequence stratigraphy and long-term paleoceanographic change in the Middle and Upper Ordovician of the eastern United States, p. 117–129. *In* B. J. Witzke, G. A. Ludvigson, and J. Day (eds.), Paleozoic sequence stratigraphy: Views from the North American Craton. Geological Society of America Special Paper 306.
- HOLLAND, S. M. AND M. E. PATZKOWSKY. 2007. Gradient ecology of a biotic invasion: Biofacies of the type Cincinnatian Series (Upper Ordovician), Cincinnati, Ohio region, USA. Palaios, 22:392–407.
- HUGHES, N. C. AND D. L. COOPER. 1999. Paleobiologic and taphonomic aspects of the "granulosa" trilobite cluster, Kope Formation (Upper Ordovician, Cincinnati region). Journal of Paleontology, 72:306–319.
- HUNDA, B. R. AND N. C. HUGHES. 2006. Evaluating paedomorphic heterochrony in trilobites: The case of the diminutive trilobite *Flexicalymene retrorsa minuens* from the Cincinnatian Series (Upper Ordovician), Cincinnati region. Evolution & Development, 9:483–498.
- HUNDA, B. R., N. C. HUGHES, AND K. W. FLESSA. 2006. Trilobite taphonomy and temporal resolution in the Mt. Orab shale bed (Upper Ordovician, Ohio, U.S.A.). Palaios, 21:26–45.
- HUNT, A. S. 1967. Growth, variation, and instar development of an agnostid trilobite. Journal of Paleontology, 41:203–208.
- KÁCHA, P. AND R. ŠARIC. 1995. Bryozoans attached to exuvia of the Ordovician trilobite *Dalmanitina* (D.) proeva. Vestnik Ceského Geologického Ustavu, 70:43–46.
- KACHA, P. AND R. ŠARIC. 2009. Host preferences in Late Ordovician (Sandbian) epibenthic bryozoans: example from the Zahořany Formation of Prague Basin. Bulletin of Geosciences, 8(1):169–178.
- KESLING, R. V. AND R. B. CHILMAN. 1975. Strata and megafossils of the Middle Devonian Silica Formation. University of Michigan Papers on Paleontology, 8, 408 p.
- KEY JR., M. M., W. B. JEFFRIES, AND H. K. VORIS. 1995. Epizoic bryozoans, sea snakes, and other nektonic substrates. Bulletin of Marine Science, 56:462–474.
- KEY JR., M. M., W. B. JEFFRIES, H. K. VORIS, AND C. M. YANG. 1996a. Epizoic bryozoans, horseshoe crabs, and other mobile benthic substrates. Bulletin of Marine Science, 58:368–384.
- KEY JR., M. M., W. B. JEFFRIES, H. K. VORIS, AND C. M. YANG. 1996b. Epizoic bryozoans and mobile ephemeral host substrata, p. 157–165. *In* D. P. Gordon, A. M. Smith, and J. A. Grant-Mackie (eds.), Bryozoans in space and time. National Institute of Water and Atmospheric Research, Wellington, New Zealand.
- KEY JR., M. M., J. W. VOLPE, W. B. JEFFRIES, AND H. K. VORIS. 1997. Barnacle fouling of the blue crab *Callinectes sapidus* at Beaufort, North Carolina. Journal of Crustacean Biology, 17:424–439.
- KEY JR., M. M. AND D. K. A. BARNES. 1999. Bryozoan colonization of the marine isopod *Glyptonotus antarcticus* at Signy Island, Antarctica. Polar Biology, 21:48–55.
- KEY JR., M. M., J. E. WINSTON, J. W. VOLPE, W. B. JEFFRIES, AND H. K. VORIS. 1999. Bryozoan fouling of the blue crab, *Callinectes sapidus*, at Beaufort, North Carolina. Bulletin of Marine Science, 64:513–533.
- KEY JR., M. M., W. B. JEFFRIES, H. K. VORIS, AND C. M. YANG. 2000. Bryozoan fouling pattern on the horseshoe crab *Tachypleus gigas* (Müller) from Singapore, p. 265–271. *In* A. Herrera C and J. B. C. Jackson (eds.), Proceedings of the 11th International Bryozoology Association Conference. Smithsonian Tropical Research Institute, Balboa, Panama.
- KITAMURA, H. AND K. HIRAYAMA. 1987. Effect of cultured diatom films on the settlement of larvae of a bryozoan *Bugula neritina*. Bulletin of the Japanese Society of Scientific Fisheries, 53:1383–1385.
- KLOC, G. J. 1992. Spine function in the odontopleurid trilobites *Leonaspis* and *Dicranurus* form the Devonian of Oklahoma. Fifth North American Paleontological Convention Abstracts and Program. Paleontological Society Special Publication, 6:167.
- KLOC, G. J. 1993. Epibionts on Selenopeltinae (Odontopleurida) trilobites. Geological Society of America Abstracts with Programs, 25(6):103.

- KLOC, G. J. 1997. Epibionts on *Dicranurus* and some related genera. Second International Trilobite Conference Abstracts with Program, St. Catharines, Ontario, p. 28.
- LESCINSKY, H. 1996. Don't overlook the epibionts. Palaios, 11:495–496.
- MARTIN, W. D. 1975. The petrology of a composite vertical section of Cincinnatian Series limestones (Upper Ordovician) of southwestern Ohio, southeastern Indiana, and northern Kentucky. Journal of Sedimentary Petrology, 45:907–925.
- MEYER, D. L. 1990. Population paleoecology and comparative taphonomy of two edrioasteroid (Echinodermata) pavements: Upper Ordovician of Kentucky and Ohio. Historical Biology, 4:155–178.
- MEYER, D. L. AND R. A. DAVIS. 2009. A sea without fish: Life in the Ordovician Sea of the Cincinnati Region. Indiana University Press, Bloomington, 346 p.
- MIKULIC, D. G. 1990. The arthropod fossil record: Biologic and taphonomic controls on its composition, p. 1–23. *In* S. J. Culver (ed.), Arthropod Paleobiology. Paleontological Society Short Course in Paleontology, 3.
- MILLER, S. A. 1875. Monograph of the class Brachiopoda of the Cincinnati Group. Cincinnati Quarterly Journal of Science, 2:6–62.
- MORRIS, R. W. AND S. H. FELTON. 1993. Symbiotic association of crinoids, platyceratid gastropods, and *Cornulites* in the Upper Ordovician (Cincinnatian) of the Cincinnati, Ohio Region. Palaios, 8:465–476.
- MORRIS, R. W. AND H. B. ROLLINS. 1971. The distribution and paleoecological interpretation of *Cornulites* in the Waynesville Formation (Upper Ordovician) of southwestern Ohio. Ohio Journal of Science, 71(3):159–170.
- NASCIMENTO, C. AND R. R. ALEXANDER. 1994. Laminar and turbulent flow patterns around models of Cambrian to Devonian trilobites in unidirectional currents: Bearing on epizoan distributions. Geological Society of America Abstracts with Programs, 26(3):65.
- NICHOLSON, H. A. 1879. On the structure and affinities of the "Tabulate Corals" of the Paleozoic Period, with critical descriptions of illustrative species. William Blackwood and Sons, Edinburgh, 342 p.
- OSGOOD, R. 1970. Trace fossils of the Cincinnati area. Palaeontographica Americana, 5:281–438.
- PECK, J. H. 1966. Upper Ordovician formations in the Maysville area, Kentucky. U.S. Geological Survey Bulletin, 1244B:1–30.
- POWERS, B. G. AND W. I. AUSICH. 1990. Epizoan associations in a Lower Mississippian paleocommunity (Borden Group, Indiana, U.S.A.). Historical Biology, 4:245–265.
- PRATT, B. R. 2001. Calcification of cyanobacterial filaments: *Girvanella* and the origin of lower Paleozoic lime mud. Geology, 29:763–766.
- PROKOP, R. 1965. Argodiscus hornyi gen. n. et sp. n. (Edrioasteroidea) from the Middle Ordovician of Bohemia and a contribution to the ecology of the edrioasteriods. Časopis Národního Muzea, Oddíl Přírodovědný, 134:30–32.RAYMOND, P. E. 1916. New and old Silurian trilobites from southeastern
- RAYMOND, P. E. 1916. New and old Silurian trilobites from southeastern Wisconsin, with notes on the genera of the Illaenidae. Bulletin of the Museum of Comparative Zoology, 60:1–41.
- RICHARDS, R. P. 1972. Autecology of Richmondian brachiopods (Late Ordovician of Indiana and Ohio). Journal of Paleontology, 46:386–405.
- Ross, D. M. 1983. Symbiotic relations, p. 163–212. *In* D. E. Bliss (ed.), The biology of Crustacea, Vol. 7. Academic Press, New York.
- SCHLOTHEIM, E. F. Von. 1820. Die Petrefactenkunde auf ihrem jetzigen Standpunkte durch die Beschreibung seiner Sammlung versteinerter und fossiler Überreste des Thier- und Pflanzenreichs der Vorwelt erläutert. Becker, Gotha, 432 p. (In German)
- SCHUMACHER, G. A. AND D. L. SHRAKE. 1997. Paleoecology and comparative taphonomy of an *Isotelus* (Trilobita) fossil lagerstätten from the Waynesville Formation (Upper Ordovician, Cincinnatian Series) of southwestern Ohio, p. 131–161. *In C. E. Brett and G. C. Baird* (eds.), Paleontological events: Stratigraphic, ecological, and evolutionary implications. Columbia University Press, New York.

- SCHUMACHER, G. A., E. M. SWINFORD, AND D. L. SHRAKE. 1991. Lithostratigraphy of the Grant Lake Limestone and Grant Lake Formation (Upper Ordovician) in southwestern Ohio. Ohio Journal of Science, 91:56–68.
- SCHUMACHER, G. A., R. C. FREY, AND S. M. BERGSTRÖM. 2000. The lithostratigraphy and biostratigraphy of the C4 sequence (Arnheim Formation) in southwestern Ohio: A test of the chronostratigraphic significance of the C5 sequence boundary. Geological Society of America Abstracts with Programs, 32(4):59.
- SEILACHER, A. 1985. Trilobite paleoecology and substrate relationships. Royal Society of Edinburgh Transactions (Earth Sciences), 76:231–237.
- SERVAIS, T., D. A. T. HARPER, A. MUNNECKEC, A. W. OWEN, AND P. M. SHEEHAN. 2009. Understanding the Great Ordovician Biodiversification Event (GOBE): Influences of paleogeography, paleoclimate, or paleoecology? GSA Today, 19:4–10.
- SHIRLEY, J. 1936. Some British trilobites of the family Calymenidae [Ordovician-Silurian] (with discussion). Quarterly Journal of the Geological Society of London, 92:384-422.
 SINGH, R. J. 1979. Trepostomatous bryozoan fauna from the Bellevue
- SINGH, R. J. 1979. Trepostomatous bryozoan fauna from the Bellevue Limestone, Upper Ordovician in the tri-state area of Ohio, Indiana and Kentucky. Bulletins of American Paleontology, 76:162–280.
- SMITH, A. M. AND C. S. NELSON. 2003. Effects of early sea-floor processes on the taphonomy of temperate shelf skeletal carbonate deposits. Earth-Science Reviews, 63:1–31.
- SNAJDR, M. 1983. Epifauna on the exuviae of Bohemian Devonian trilobites. Casopis pro Mineralogii a Geologii, 28:181–186.
- TAPANILA, L. 2005. Palaeoecology and diversity of endosymbionts in Palaeozoic marine invertebrates: trace fossil evidence. Lethaia, 38:89–99.
- TAYLOR, P. D. 1990. Encrustors, p. 346–351. In D. E. G. Briggs and P. R. Crowther (eds.), Palaeobiology: A synthesis. Blackwell Scientific, Oxford.
- TAYLOR, P. D. AND M. A. WILSON. 2002. A new terminology for marine organisms inhabiting hard substrates. Palaios, 17:522–525.
- TAYLOR, P. D. AND M. A. WILSON. 2003. Palaeocology and evolution of marine hard substrate communities. Earth-Science Reviews, 62:1–103.
- TAYLOR, W. L. AND C. E. BRETT. 1996. Taphonomy and paleoecology of echinoderm lagerstätten from the Silurian (Wenlockian) Rochester Shale. Palaios, 11:118–140.
- TETREAULT, D. K. 1992. Paleoecologic implications of epibionts on the Silurian lichid trilobite Arctinurus, p. 289. In S. Lidgard and P. R. Crane (eds.), North American Paleontological Convention Abstracts and Program. Paleontological Society, Special Publication 6. TETREAULT, D. K. 1997. Trilobite behavior in the Silurian. Second
- TETREAULT, D. K. 1997. Trilobite behavior in the Silurian. Second International Trilobite Conference, Brock University, St. Catharines, Ontario. Abstracts with Programs. p. 48–49.
- ULRICH, E. O. 1882. American Palaeozoic Bryozoa. Journal of the Cincinnati Society of Natural History, 5:121–175, 233–257.
- VANCE, R. R. 1978. A mutualistic interaction between a sessile marine clam and its epibionts. Ecology, 59:679–685.
- VOGEL, S. 1981. Life in moving fluids. Willard Grant Press, Boston, 352 p.
- WAUGH, D. A., R. M. FELDMANN, R. S. CRAWFORD, S. L. JAKOBSEN, AND K. B. THOMAS. 2004. Epibiont preservational and observational bias in fossil marine decapods. Journal of Paleontology, 78:961–972.
- WEIR, G. W., W. L. PETERSON, AND W. C. SWADLEY. 1980. Measured sections of Ordovician strata in Indiana and Ohio. U.S. Geological Survey Open-File Report, 80–235, 81 p.
- WYSE JACKSON, P. N. AND M. M. KEY JR. 2007. Borings in trepostome bryozoans from the Ordovician of Estonia: Two ichnogenera produced by a single maker, a case of host morphology control. Lethaia, 40:237– 252.

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